

Inhibition of pancreatic cancer stem cells by *Rauwolfia vomitoria* extract

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Abstract. The poor treatment outcomes of pancreatic cancer are linked to an enrichment of cancer stem cells (CSCs) in these tumors, which are resistant to chemotherapy and promote metastasis and tumor recurrence. The present study investigated an extract from the root of the medicinal plant *Rauwolfia vomitoria* (Rau) for its activity against pancreatic CSCs. *In vitro* tumor spheroid formation and CSC markers were tested, and *in vivo* tumorigenicity was evaluated in nude mice. Rau inhibited the overall proliferation of human pancreatic cancer cell lines with a 50% inhibitory concentration (IC₅₀) ranging between 125 and 325 µg/ml, and showed limited cytotoxicity towards normal epithelial cells. The pancreatic CSC population, identified using cell surface markers or a tumor spheroid formation assay, was significantly reduced, with an IC₅₀ value of ~100 µg/ml treatment for 48 h and ~27 µg/ml for long-term tumor spheroid formation. The levels of CSC-related gene Nanog and nuclear β-catenin were decreased, suggesting suppression of the Wnt/β-catenin signaling pathway. *In vivo*, 20 mg/kg of Rau administered five times per week by oral gavage significantly reduced the tumorigenicity of PANC-1 cells in immunocompromised mice. Taken together, these data showed that Rau preferentially inhibited pancreatic cancer stem cells. Further investigation is warranted to examine the potential of Rau as a novel treatment for pancreatic cancer.

Introduction

With an estimation of 55,440 new cases (29,200 in men, 26,240 in women), and a mortality rate of 44,330 (23,020 men, 21,310 women) in 2018, pancreatic cancer remains the fourth leading cause of cancer-associated mortality in the USA,

and is one of the most life-threatening malignancies (1). As pancreatic cancer shows high level of heterogeneity and often metastasizes early, management of pancreatic cancer has always been challenging (2,3). The majority of patients with pancreatic cancer (~53%) are diagnosed at an advanced stage, for whom the 5-year-survival rate is only 2-5%, which is among the lowest of all types and stages of malignancy (4). Even in the 10% patients who are diagnosed at early stages, the 5-year-survival rate is only 32%. Gemcitabine as the first line chemotherapy provides limited benefit on the overall survival rate of patients with locally advanced and metastatic pancreatic cancer (5,6). Numerous efforts have been made to improve the treatment outcome. The development of treatment regimens, including FOLFIRINOX (7,8) or nab-paclitaxel plus gemcitabine (9) have led to improvement in survival and response rates, however, they significantly increase toxic side effects (10,11). Novel treatment options are urgently required for pancreatic cancer.

One of the reasons for the poor treatment outcomes is that pancreatic cancer has an enriched cancer stem cell (CSC) population (6). CSCs are responsible for tumor generation (3), are resistant to current chemotherapy and radiation therapies (12), and are prone to metastasis (13). The cells survive current treatments and eventually give rise to new tumors either at the primary or metastatic sites (14-16). Depending on the microenvironment, a CSC can be characteristically quiescent, and the dormancy protects them from chemotherapeutic agents that target actively dividing cells (17). Alternately, a CSC can divide and generate daughter cells which give rise to all cell types found in a particular bulk of tumor (18), and/or generate daughter cells which do not differentiate but maintain the full potential for differentiation as the parent stem cell (self-renewal) (17). The self-renewal ability maintains the number of CSCs within the tumor, whereas its descendent progeny constitute the bulk of the tumor. CSCs also exhibit unique features, including drug resistance and metastatic ability. If a treatment does not eliminate CSCs, the CSCs eventually promote tumor recurrence. Therefore, therapies that inhibit CSCs offers promise in eliminating the whole cancer cell population.

Herbal preparations of *Rauwolfia vomitoria* (Rau), a tropical shrub in the family Apocynaceae, is a traditional folk medicine in Africa used to treat a variety of conditions, including hypertension (19,20), fever (21,22), gastrointestinal

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diseases (23), liver diseases (24) and cancer (25). The extract as a whole mixture is widely used as a health supplement. Extracts from the root bark of this plant are enriched with β -carboline alkaloids and indole alkaloids (26). β -carboline alkaloids have been reported to have several bioactivities, including antitumor effects (27,28). In our previous study, it was reported that an extract of Rau, with its hypotensive component reserpine removed, induced pancreatic cancer cell apoptosis, and inhibited pancreatic tumor growth in mice (29). The combination of Rau and gemcitabine showed synergistic antitumor effects (29). In the present study, the activities of the same extract on inhibiting pancreatic CSCs *in vitro* and *in vivo* were investigated.

Materials and methods

Cell lines and reagents. The PANC-1, AsPC-1, HPAF-II, BxPC-3 and Mia PaCa-2 human pancreatic cancer cell lines were obtained from the American Type Culture Collection (ATCC, Manassas, VA, USA) and maintained in the laboratory. The MRC-5 immortalized human lung epithelial cell line was provided by Dr Sitta Sittampalam at the National Center for Advancing Translational Sciences, NIH (Bethesda, MD, USA), and was used as a comparison to the cancer cells. All cells were cultured at 37°C in 5% CO₂/95% air in recommended growth media: PANC-1 and Mia PaCa-2 in DMEM (cat no. 10-013-CV; Corning, Inc., Corning, NY, USA); AsPC-1 and BxPc-3 in RPMI-1640 (cat. no. 10-040-CV; Corning, Inc.) and HPAF-II in EMEM (cat. no. 10-010-CV; Corning Inc.), containing 10% fetal bovine serum (FBS; Sigma-Aldrich, St. Louis, MO, USA; cat. no. F0926) and 1% antibiotics (cat. no. 30-001-C; Corning, Inc.). The Rau extract was provided by Natural Source International, Ltd. (New York, NY, USA) and was prepared in sterile phosphate-buffered saline (PBS) in 10 mg/ml stock solutions and stored at -20°C.

Cell viability assay. The cells were assessed for viability using a 3-(4, 5-dimethylthiazol-2-yl)-2, 5-diphenyltetrazolium bromide (MTT) assay at 48 h of treatment. Cells in the exponential growth phase were exposed to serial dilutions of Rau for 48 h. The medium was then replaced with fresh media containing MTT and cells were incubated for 4 h at 37°C. The colorimetric MTT assay assesses relative proliferation, based on the ability of living, but not dead cells, to reduce MTT to formazan. The cells did not reach a plateau phase during the incubation period. The 50% inhibitory concentration (IC₅₀) was defined as the concentration of drug that inhibited cell growth by 50% relative to the untreated control. Pilot experiments for each cell line were performed to optimize cell density and assay duration, and to center drug dilution series approximately on the IC₅₀.

Tumor spheroid formation assay. For the PANC-1 cells, a single-cell suspension was plated into 24-well ultra-low attachment plates (Corning Inc.) at a density of 5,000 cells/well in stem cell media and incubated at 37°C in a humidified atmosphere of 95% air and 5% CO₂. For the MIA PaCa-2 cells, a single-cell suspension was plated into 96-well ultra-low attachment plates (Corning Inc.) at a density of 100 cells/well in stem cell media and incubated under the same conditions. The stem cell

media consisted of DMEM (Corning Inc.) supplemented with 1X B27 Supplement, 20 ng/ml human basic fibroblast growth factor, 20 ng/ml epidermal growth factor, 100 U/ml penicillin/streptomycin (Invitrogen; Thermo Fisher Scientific, Inc.) and 4 μ g/ml heparin calcium salt (Thermo Fisher Scientific, Inc.). The PANC-1 spheroids were counted following 4 weeks of culture and the MIA PaCa-2 spheroids were counted following 2 weeks of culture under the microscope. Spheroid diameter was measured using ImageJ software v1.48 (NIH, Bethesda, MD, USA).

Flow cytometry for the detection of CSC surface markers. Rau has marked autofluorescence in two ranges of emission wavelength, at 400-600 nm and 800-900 nm, overlapping the emission wavelength of several fluorescent labeling molecules. Therefore, PE-Cy7-conjugated CD24 and APC-conjugated EpCam antibodies were used as indicative markers for pancreatic CSCs (CD24⁺EpCam⁺) to avoid overlapping with Rau autofluorescence. The cells were exposed to various concentrations of Rau for 24 or 48 h. The cells were then washed with PBS three times, and resuspended in binding buffer (PBS supplemented with 0.1% bovine serum albumin (BSA; Fisher BioReagents, Waltham, MA, USA; cat. no. BP1605-100) for 15 min. PE-Cy-7-conjugated anti-CD24 antibody (dilution 1:100; cat. no. 311119; BioLegend, Inc., San Diego, CA, USA) and APC-conjugated anti-EpCam antibody (dilution 1:100; cat. no. 324207; BioLegend, Inc.) were added into the cell suspension and incubated for 15 min according to the manufacturer's protocol. The cells were washed in PBS three times following staining and then analyzed using a BD LSR II flow cytometer. The data were normalized to cell death, as follows: Normalized CSC population = original CSC population detected with flow cytometry x % cell viability detected with the MTT assay.

Flow cytometry for sorting of side population from pancreatic cancer cells. Dye Cycle Violet (DCV; Invitrogen; Thermo Fisher Scientific, Inc.) was used for staining of the non-CSC population. Cells that efficiently exclude DCV from the cytoplasm are considered CSC-like population (DCV⁻ cells). The MIA PaCa-2 cells were suspended at a density of 1x10⁶ cells/ml in DME supplemented with 10% FBS and 10 mM HEPES. DCV (10 μ M) was added and incubated for 30 min at room temperature. The cells were then washed twice with PBS, and resuspended in DMEM supplemented with 10% FBS and 10 mM HEPES for 1 h. The cells were transferred to ice-cold HBSS/2% FBS/10 mM HEPES buffer immediately prior to flow cytometric sorting. The DCV⁻ and DCV⁺ cells were separately collected for further analysis. Gate setting was performed using cells treated with a pump inhibitor (verapamil; 200 μ M) prior to DCV staining.

SDS PAGE and western blot analysis. The cells were lysed with RIPA buffer containing protease inhibitors and phosphatase inhibitors (Sigma-Aldrich; EMD Millipore) followed by sonication for 10 sec. Either whole cell lysate or supernatant was used for further experiments, depending on the proteins of interest. The BCA method was used for protein quantification (Pierce BCA protein assay kit; Thermo Fisher Scientific, Inc.). SDS-PAGE and western blot analyses were performed

as routine: 10 μ g total protein or 2 μ g nuclear or cytoplasmic fractions were loaded on a 10% SDS-PAGE gel, and electrophoresis was performed at 60 V for 35 min followed by 90 V for 90 min. Proteins were transferred to PVDF membrane (cat.no.ISEQ00010;EDMillipore,Burlington,USA) overnight. Membrane was then blocked with 5% blocking grade blocker (cat. no. 170-6404; Bio-Rad Laboratories, Inc., Hercules, CA, USA) in 1X TBS-T (Tween-20, 0.1%) for 2 h at room temperature with constant shaking. Primary and secondary antibodies were from Cell Signaling Technology Inc. (Danvers, MA, USA): rabbit anti- β -catenin (dilution 1:1,000; cat. no. 9582), rabbit anti-vinculin (dilution 1:1,000; cat. no. 4650), rabbit anti-Histone H3 (dilution 1:2,000; cat. no. 4499), rabbit anti-Nanog (dilution 1:2,000; cat. no. 4903), mouse anti- β -actin (dilution 1:2,000; cat. no. 3700), and goat anti-rabbit (dilution 1:5,000; cat. no. 7074) or anti-mouse (dilution 1:5,000; cat. no. 7076) IgG. Primary antibodies were incubated overnight at 4°C and secondary antibodies were incubated for 2 h at room temperature. The blots were established using a chemiluminescence detection kit (Pierce ECL; cat. no. 32106) or ECL Plus Western Blotting Substrate (cat. no. 32132) which were both from Thermo Fisher Scientific, Inc.

RNA isolation, cDNA synthesis and reverse transcription-quantitative polymerase chain reaction (RT-qPCR) analysis. Total RNA was extracted from cells or tissue samples using TRIzol reagent according to the protocol of the manufacturer (Invitrogen; Thermo Fisher Scientific, Inc.). cDNA synthesis was performed with 1 μ g of total RNA using an Omniscript RT kit according to the manufacturer's protocol (Qiagen, Inc., Valencia, CA, USA). cDNA was diluted 1:5 in DEPC-treated nanopure water and used for further analysis. RT-qPCR analysis was performed using a Bio-Rad iQ iCycler detection system with iQ SYBR green supermix (Bio-Rad Laboratories, Inc.). The reactions were performed in a total volume of 10 μ l, including 5 μ l of 2X iQ SYBR Green supermix, 0.4 μ l of primers at 20 pmol/ μ l and 0.4 μ l of cDNA template. Primer sequences are as follows: BCL2L2 (F, GCGGAGTTCACAGCTCTATAC and R, AAAAGGCCCTACAGTTACCA); Cox-2 (F, CTGGCGCTCAGCCATACAG and R, CGCAC TTACTGGTCAAATCCC); MMP14 (F, GGCTACAGC AATATGGCTACC and R, GATGGCCGCTGAGAGTGAC); MYC (F, TCCCTCCACTCGGAAGGAC and R, CTGGT GCATTTTCGGTTGTTG); hBim (F, TACTCCAGTGC AGTCTCCTC and R, TCCCATCTTTCCTAACACAG); hDppa4 (F, AAAAGCAAGAAGGGAGAGTGA and R, CGG AGATTGCACTGAACTGA); hEsrrb (F, TCAGAGAGCA GCCATACCT and R, GCGTCACAACT CCTCCTTC); hOct4 (F, GAGAATTTGTTCTCAGTGC and R, GTTCCC AATTCCTTCC TTAGTG); hSox2 (F, ATGGGTTTCG GTGGTCAAGT and R, GTGGATGGGATTGGTGTCTC); hTbx3 (F, GAAGAAGAGGTGGAGGACGA and R, ATTCAG TTTCGGGGAACAAG); hTcl1 (F, GATACCGATCCTCA GACTCCA and R, GAGGACAGAAGGGACAGAA); GAPDH (F, CCAGGTGGTCTCCTCTGACTTCAACA and R, AGGGTCTCTCTTCTCCTTGTGCTC). All qPCR were run according to the following thermocyclers: Initial denaturation and enzyme activation at 95°C for 3 min, followed by 40 cycles of denaturing (95°C for 15 sec), annealing (55-60°C for 30 sec), and extension (72°C for 30 sec). Melt

curve was carried out at 55-95°C (in 0.5°C increments) for 30 sec.

All reactions were performed in four repeats for every sample and with three independent experiments for each group. GAPDH was used as housekeeping gene for normalization. Gene expression was quantified using $\Delta\Delta$ CT method and $2^{-\Delta\Delta C_t}$ was used as the relative expression changes for each gene (30).

Pancreatic cancer xenograft mouse model. All animal experiments followed a protocol approved by the Institutional Animal Care and Use Committee of the University of Kansas Medical Center (Kansas City, KS, USA). Single treatment and repeated treatments were each used for the measurement of tumorigenicity. In the single treatment model, the PANC-1 pancreatic cancer cells at three densities were used for tumor inoculation (2×10^4 cells per injection, 2×10^5 cells per injection, or 1×10^6 cells per injection). The PANC-1 cells were suspended in PBS as single cell suspension and then mixed with either 200 mg/ml Rau or PBS. At each cell injection number, cells mixed with Rau were injected subcutaneously into the left flank of the mouse, and cells mixed with PBS were injected into the right flank of the same mouse. A total of 10 mice (Athymic Ncr nu/nu, female, 4-6 weeks, 15-20 g) were used for each cell density. The formation of tumors were monitored daily, and longitudinal tumor growth was measured using calipers. Mice were housed in 5 mice/cage in a sterile rodent room with 12-h/12 h-light/dark cycle. Housing was handled by the University of Kansas Medical Center Laboratory Animal Resources following standard protocol.

In the repeated treatment model, a single cell suspension of PANC-1 cells was mixed with 200 mg/ml Rau, and then inoculated into 10 mice at 2×10^5 cells per injection, in the left and right flanks. Treatment was started the following day with oral gavage of 20 mg/kg Rau, five times per week for 3 weeks. In the control group (10 mice), the mice were inoculated with the same number of cells in PBS, and were then gavaged with an equivalent volume of saline solution. Tumor formation was monitored daily, and longitudinal tumor growth was measured using calipers.

Statistical analysis. Statistical analysis was performed using IBM SPSS statistical software, version 23 (IBM PSS, Armonk, NY, USA). Student's t-test and a log-rank test were used. $P < 0.05$ was considered to indicate a statistically significant difference.

Results

Inhibition of pancreatic cancer tumor spheroid formation in vitro. Five human pancreatic cancer cell lines (PANC-1, MiA PaCa-2, AsPC-1, HPAF-II and BxPC-3) and an immortalized epithelial cell line (MRC-5) were treated with various concentrations of Rau, and cell viability was detected 48 h later. Rau inhibited the proliferation of all five cancer cells (Fig. 1A), with IC_{50} values ranging between 140 and 317 μ g/ml. The non-cancerous MRC-5 epithelial cell line was less affected by Rau treatment, with a higher IC_{50} value of 567 μ g/ml (Fig. 1B). These results are consistent with our previous findings that Rau inhibited the overall proliferation of pancreatic cancer cells (29).

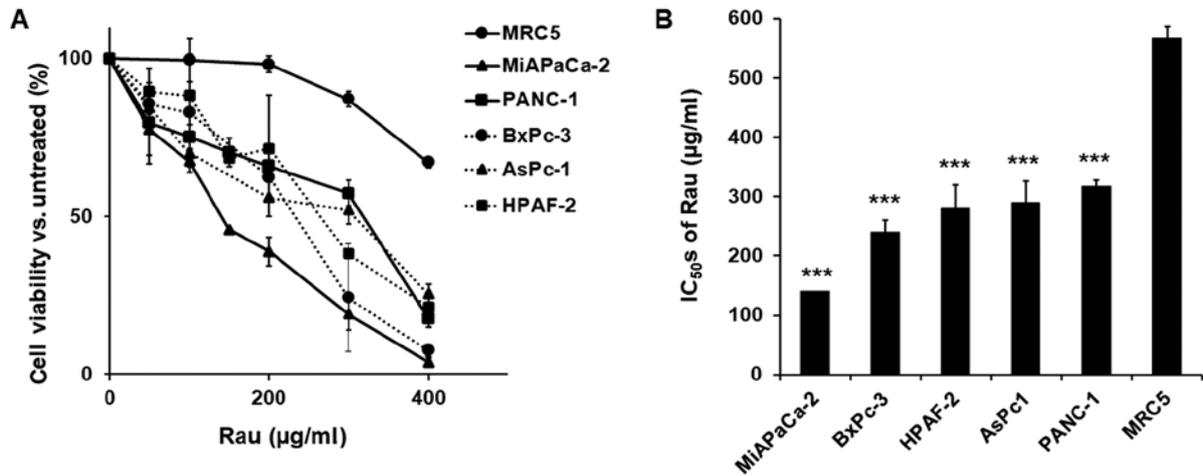


Figure 1. Inhibition of the proliferation of pancreatic cancer cells by Rau. (A) Dose-response curves. Human PANC-1, AsPC-1, HPAF-II, BxPC-3 and MiA PaCa-2 pancreatic cancer cells were exposed to serial concentrations of Rau for 48 h. Cell viability was detected using a 3-(4, 5-dimethylthiazol-2-yl)-2, 5-diphenyltetrazolium bromide assay. An immortalized non-cancerous MCR-5 epithelial cell line was subjected to the same treatment. (B) IC₅₀ values of Rau in pancreatic cancer cells and MRC-5 cells. ***P<0.001, compared with the IC₅₀ of MRC-5 cells. All values are expressed as the mean ± standard deviation of three independent experiments each performed in triplicate. Rau, *Rauwolfia vomitoria*; IC₅₀, 50% inhibitory concentration.

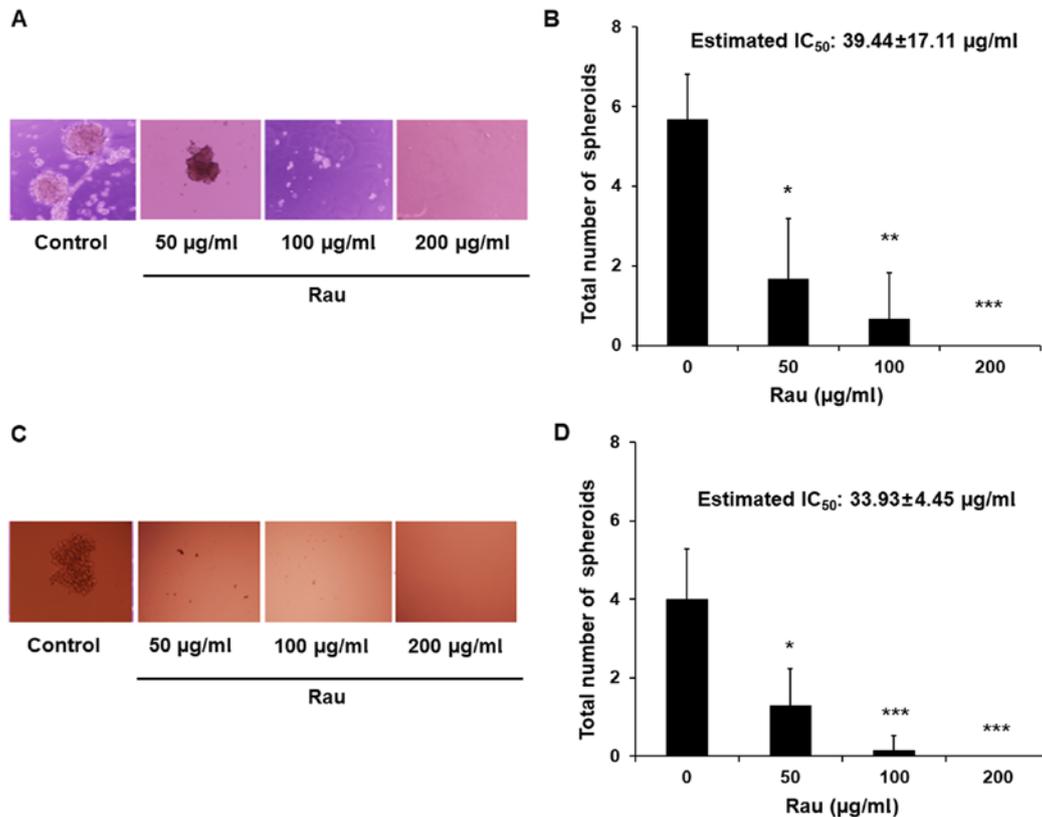


Figure 2. Inhibition of pancreatic tumor spheroids by Rau. (A) Representative images of the spheroids with and without Rau treatment (magnification, x10). A PANC-1 single-cell suspension was plated into 24-well ultra-low attachment plates at a density of 5,000 cells/well in stem cell media. Tumor spheroids were counted 4 weeks later. (B) Number of PANC-1 spheroids (mean ± standard deviation of three independent experiments). *P<0.05, **P<0.01 and ***P<0.001, compared with the untreated control. (C) Representative images of the MIA PaCa-2 spheroids with and without Rau treatment. MIA PaCa-2 single-cell suspension was plated into 96-well ultra-low attachment plates at a density of 100 cells/well in stem cell media. Tumor spheroids were counted 2 weeks later. (D) Number of MIA PaCa-2 spheroids (mean ± standard deviation of three independent experiments). Rau, *Rauwolfia vomitoria*; IC₅₀, 50% inhibitory concentration.

To investigate the inhibitory effect of Rau in CSCs, a tumor spheroid formation assay was performed. The ability to form tumor spheroids is an *in vitro* indication of the tumorigenic capacity and self-renew ability of CSCs. When cancer cells

are cultured in non-adherent, serum-free conditions, non-CSC populations die by anoikis, whereas CSCs overcome anoikis and go through division leading to the formation of tumor spheroids (32,33). Single cell suspensions were treated with

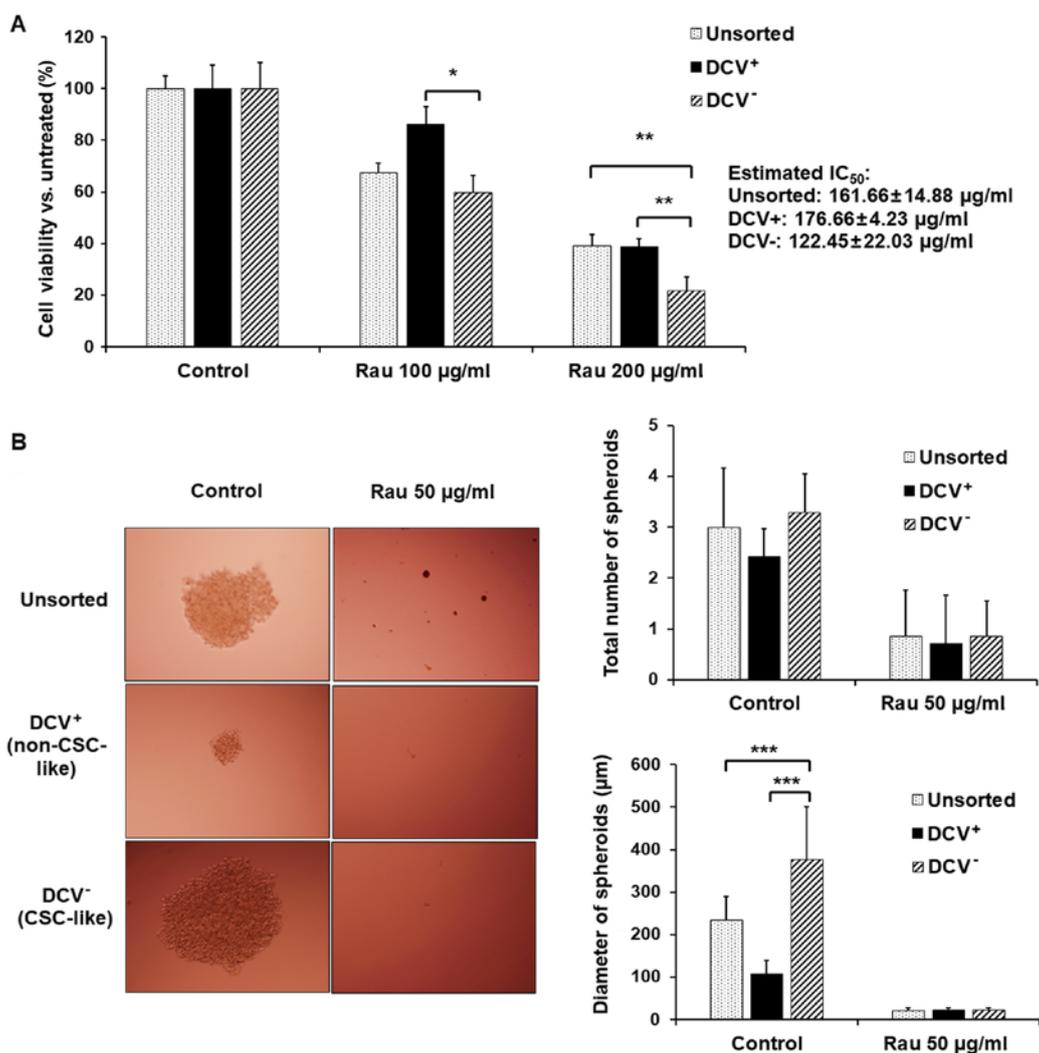


Figure 3. Inhibition of pancreatic CSC-like subpopulations by Rau. (A) Cell proliferation of unsorted cells, DCV⁺ cells (non-CSC-like) and DCV⁻ cells (CSC-like) with Rau treatment for 48 h (mean ± standard deviation of three independent experiments). (B) Representative images of the MIA PaCa-2 spheroids from unsorted cells, DCV⁺ cells and DCV⁻ cells with and without Rau treatment (magnification, x10). The number and size of MIA PaCa-2 spheroids are shown in bar graph. *P<0.05; **P<0.01 and ***P<0.001. CSC, cancer stem cell; Rau, *Rauwolfia vomitoria*; DCV, Dye Cycle Violet; IC₅₀, 50% inhibitory concentration.

Rau and tumor spheroids were counted 4 weeks later. Data showed that Rau significantly reduced the number of the PANC-1 tumor spheroids at the concentrations of 50 and 100 µg/ml, and completely eliminated the tumor spheroids at 200 µg/ml (Fig. 2A and B). The estimated IC₅₀ value for tumor spheroids inhibition is 39.44 µg/ml. By contrast, the IC₅₀ value of Rau to the bulk of PANC-1 cells was 317 µg/ml (Fig. 1B). The MIA PaCa-2 pancreatic cancer cells were also treated by Rau for the detection of tumor spheroids. Similar results were obtained. Rau reduced the number of the MIA PaCa-2 spheroids at 50 µg/ml, and completely inhibited spheroid formation at ≥100 µg/ml (Fig. 2C and D). The estimated IC₅₀ value of 34 µg/ml (Fig. 2D) was lower than the IC₅₀ value for the bulk of the MIA PaCa-2 cells (Fig. 1A).

Cells with stemness features are reported to exclude dyes as side populations (34,35). In order to separate the CSC-like population, MIA PaCa-2 cells were sorted using flow cytometry with DCV staining. The DCV⁻ cells (CSC-like) and DCV⁺ (non CSC-like) cells were collected and treated with Rau. Rau inhibited the viability in all unsorted, DCV⁺ and DCV⁻ cells,

preferentially inhibiting DCV⁻ cells (Fig. 3A). The estimated IC₅₀ values were 162 µg/ml in unsorted cells, 177 µg/ml in DCV⁺ cells and 122 µg/ml in DCV⁻ cells. This result suggested that Rau preferentially inhibited CSC-like cells.

Tumor spheroid formation was detected. Although cell spheroids were also formed in the DCV⁺ cell culture, they were significantly smaller (Fig. 3B). By contrast, DCV⁻ cells formed large spheroids, as expected. As there is currently no exclusive way to pin-point pancreatic CSCs, the formation of spheroids in DCV⁺ cells may due to the remaining CSC-like cells in the DCV⁺ population. However, the DCV staining and sorting provided a side population enriched with 'stemness'. Rau at 50 µg/ml inhibited spheroids in the DCV⁻ and DCV⁺ populations (Fig. 3B), a result consistent with those in unsorted cells.

Reduction of the CSC marker-positive cell population. The effects of Rau on CSCs in a shorter time period were also examined. The PANC-1 cells were treated with Rau for 24 or 48 h at concentrations of 50, 100 or 200 µg/ml. The pancreatic

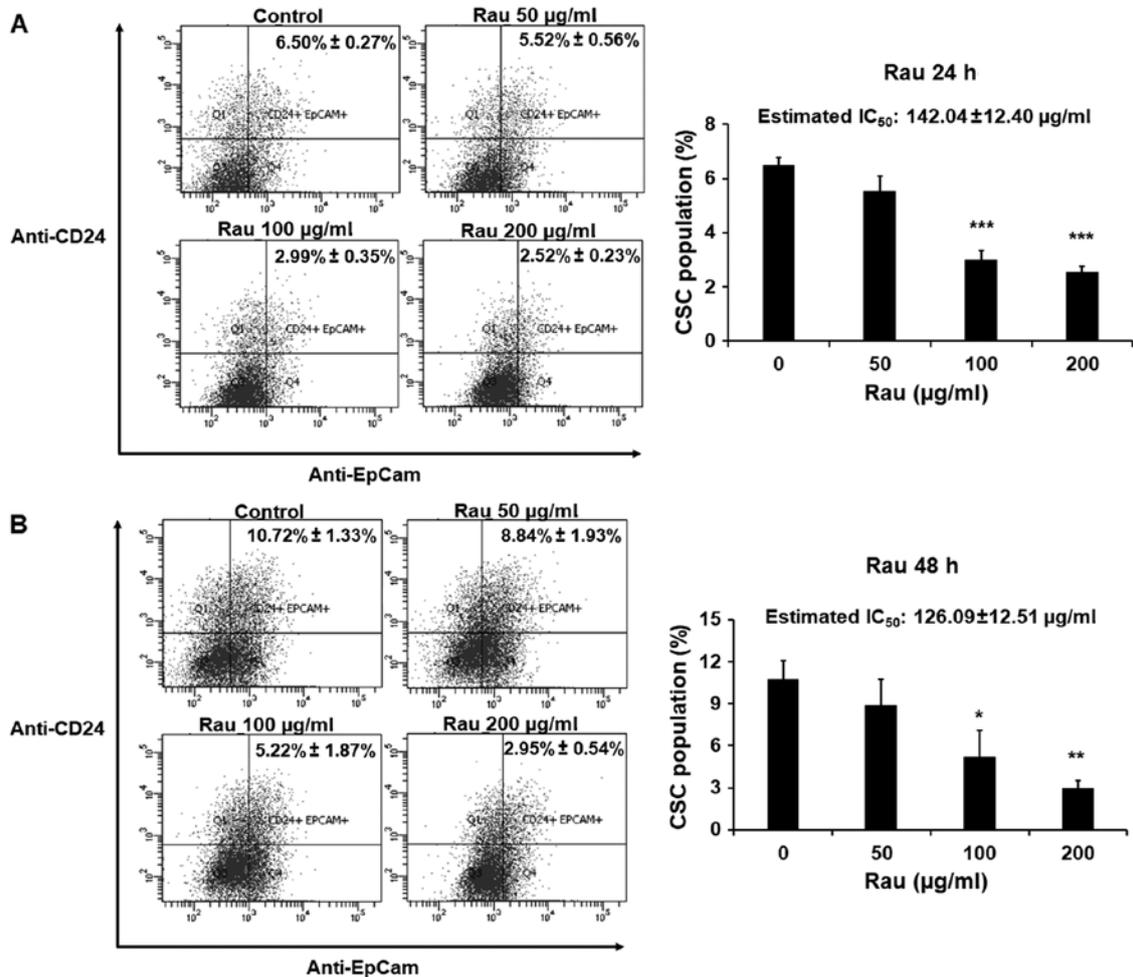


Figure 4. Inhibition of CSC populations by Rau. PANC-1 cells were treated with Rau for (A) 24 and (B) 48 h at the indicated concentrations. Cells were then stained with fluorescent conjugated antibodies for CD24 and EpCam, followed by flow cytometric analysis. Left panels show the EpCam (APC) and CD24 (PE) positive cells. The percentages of CD24⁺EpCam⁺ cells were quantified and shown in the bar graph (mean \pm standard deviation of three experiments). The data were normalized to cell death. *P<0.05; **P<0.01; ***P<0.001, compared with the untreated group. CSC, cancer stem cell; Rau, *Rauwolfia vomitoria*; IC₅₀, 50% inhibitory concentration.

CSC markers CD24 and EpCAM were examined by immune staining followed by flow cytometric analysis. Rau reduced the CD24⁺EpCam⁺ cell population following 24 and 48 h treatment (Fig. 4A and B). In the control groups, CD24⁺EpCam⁺ cells consisted of 6.50-10.72% of the whole population. At the concentration of 200 $\mu\text{g/ml}$, Rau significantly reduced CD24⁺EpCam⁺ cells to 2.52-2.95% following 24 and 48 h of treatment (Fig. 4A and B). At a lower concentration of 100 $\mu\text{g/ml}$, Rau also significantly reduced the CD24⁺EpCam⁺ cells to 2.99-5.22% following 24 and 48 h of treatment (Fig. 4B). It was estimated that the IC₅₀ value at 24-h treatment was 142.04 \pm 12.40 $\mu\text{g/ml}$, and at 48 h treatment was 126.09 \pm 12.51 $\mu\text{g/ml}$ (Fig. 4A and B), and these values were lower than the IC₅₀ values for the bulk tumor cells. These data are consistent with the results above showing that Rau preferentially inhibited pancreatic CSCs.

One of the essential pathways in maintaining the self-renewal and spheroid formation capacities of CSCs is activation of the canonical Wnt/ β -catenin signaling pathway (18,36). When there is active Wnt signaling, the β -catenin degradation complex in the cytosol dissociates, and β -catenin accumulates in the nucleus and functions as a transcriptional factor

to upregulate genes that promote CSC stemness, including Nanog (37). In the present study, the cytoplasmic and nuclear fractions of the PANC-1 cells were each examined for β -catenin levels with or without Rau treatment. Treatment with Rau (100 $\mu\text{g/ml}$) for 24 and 48 h reduced the levels of β -catenin in the nucleus (Fig. 5A), whereas the cytoplasmic β -catenin levels were not changed (Fig. 5A). A panel of β -catenin downstream target genes, including B-cell lymphoma 2-like 2 (BCL2L2), cyclooxygenase-2 (COX-2), matrix metalloproteinase (MMP)14 and MYC, were examined by RT-qPCR analysis (Fig. 5B). Following treatment for 48 h, the expression of MYC was significantly decreased by Rau treatment, which is consistent with Wnt/ β -catenin signaling pathway inhibition. Studies have shown that the stem cell-related gene Nanog has the ability to induce β -catenin phosphorylation and enhances its degradation (38). Therefore, the present study examined the expression of Nanog by western blot analysis. Nanog was increased following 24 h of Rau treatment, and was then decreased following 48 h of Rau treatment (Fig. 5C). It was hypothesized that the increase in Nanog at the earlier time point enhanced β -catenin degradation and therefore suppressed nuclear levels of β -catenin. The suppressed β -catenin levels subsequently

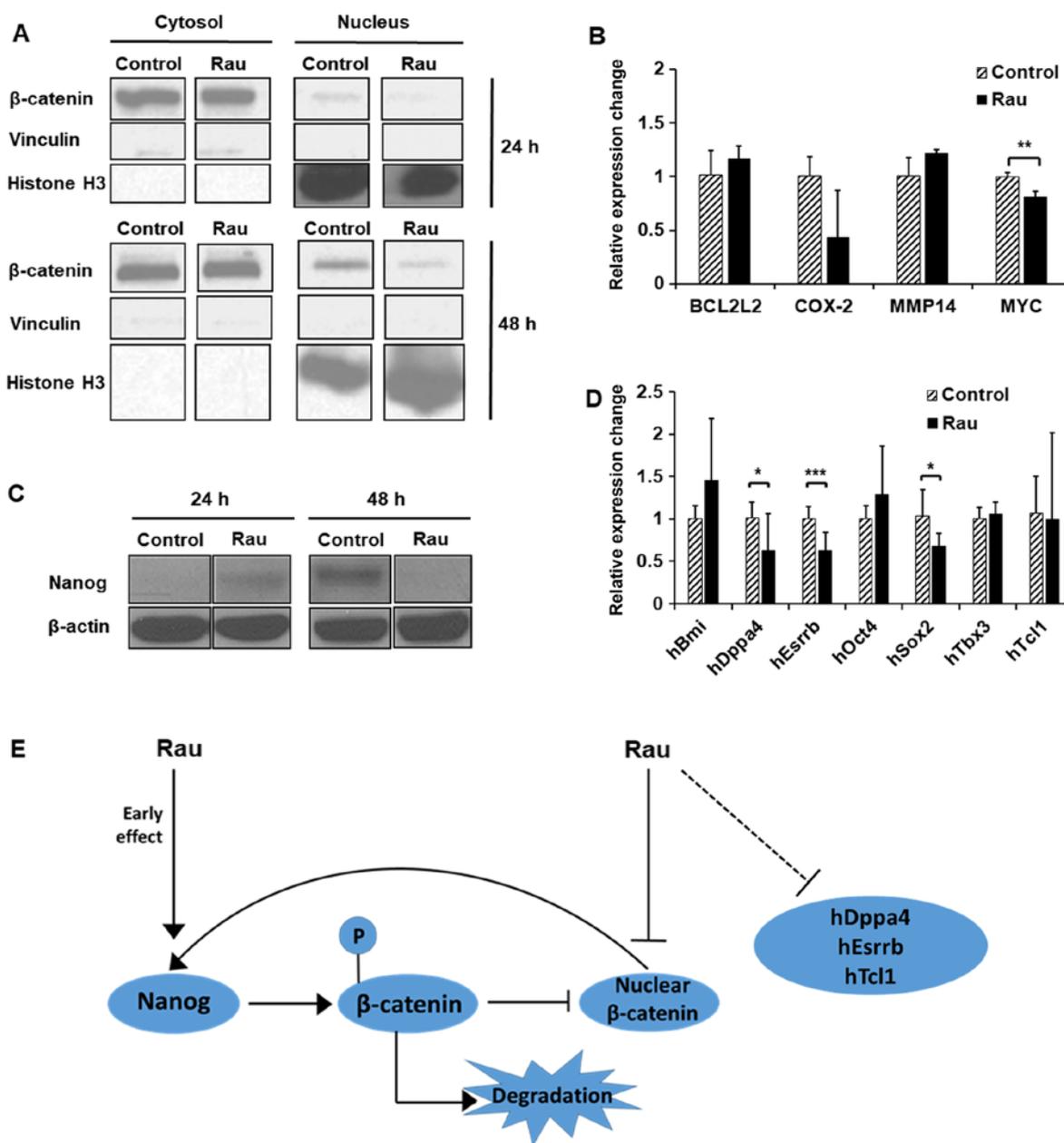


Figure 5. Decrease of nuclear β -catenin by Rau. PANC-1 cells were treated with Rau at 100 $\mu\text{g/ml}$ for 24 and 48 h. (A) Expression of β -catenin was detected by western blot analysis in cytoplasmic and nuclear fractions. Vinculin was a loading control for cytoplasmic proteins, and histone H3 was a loading control indicative for the nuclear fraction. (B) Expression of β -catenin downstream target genes at 48 h of Rau treatment, detected by RT-qPCR analysis. (C) Expression of Nanog was detected by western blot analysis. (D) Expression of CSC-related genes following 48 h of Rau treatment, detected by RT-qPCR analysis. (E) Suggested mechanism of Rau inhibiting Nanog and nuclear β -catenin. Rau treatment has an early effect in increasing the expression of Nanog, which leads to the phosphorylation and degradation of β -catenin, which represses nuclear β -catenin. The decreasing level of nuclear β -catenin negatively influenced the expression of Nanog. Rau treatment also appeared to directly inhibit β -catenin nuclear accumulation. Both can result in overall suppression of levels of Nanog and nuclear β -catenin. Rau also inhibited the RNA level of CSC-related genes, including Dppa4 and Esrrb. * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$, compared with the untreated control group. Rau, *Rauwolfia vomitoria*; RT-qPCR, reverse transcription-quantitative polymerase chain reaction; BCL2L2, B-cell lymphoma 2-like 2, COX-2, cyclooxygenase-2, MMP14, matrix metalloproteinase 14; Dppa4, developmental pluripotency associated 4, Esrrb, estrogen related receptor β ; Oct4, octamer-binding transcription factor 4; Tbx3, T-box 3; Tcl1, T cell leukemia 1.

resulted in inhibition of the expression of Nanog at a later time-point (39,40). As a result, the Nanog and Wnt signaling pathway was suppressed by Rau.

A panel of other CSC-related genes were also examined by RT-qPCR analysis (31). Data showed that the expression levels of developmental pluripotency associated 4 (Dppa4), estrogen related receptor β (Esrrb) and SRY-box 2 (Sox2) were inhibited following 48 h of Rau treatment (Fig. 5D).

Taken together, Rau treatment had an early effect in increasing the expression of Nanog, which led to the phosphorylation and degradation of β -catenin, and repressed the nuclear level of β -catenin. The decreasing level of nuclear β -catenin negatively affected the expression of Nanog. Rau treatment also appeared to directly inhibit the nuclear accumulation of β -catenin and other CSC-related genes, including MYC, resulting in the overall suppression of Nanog and nuclear

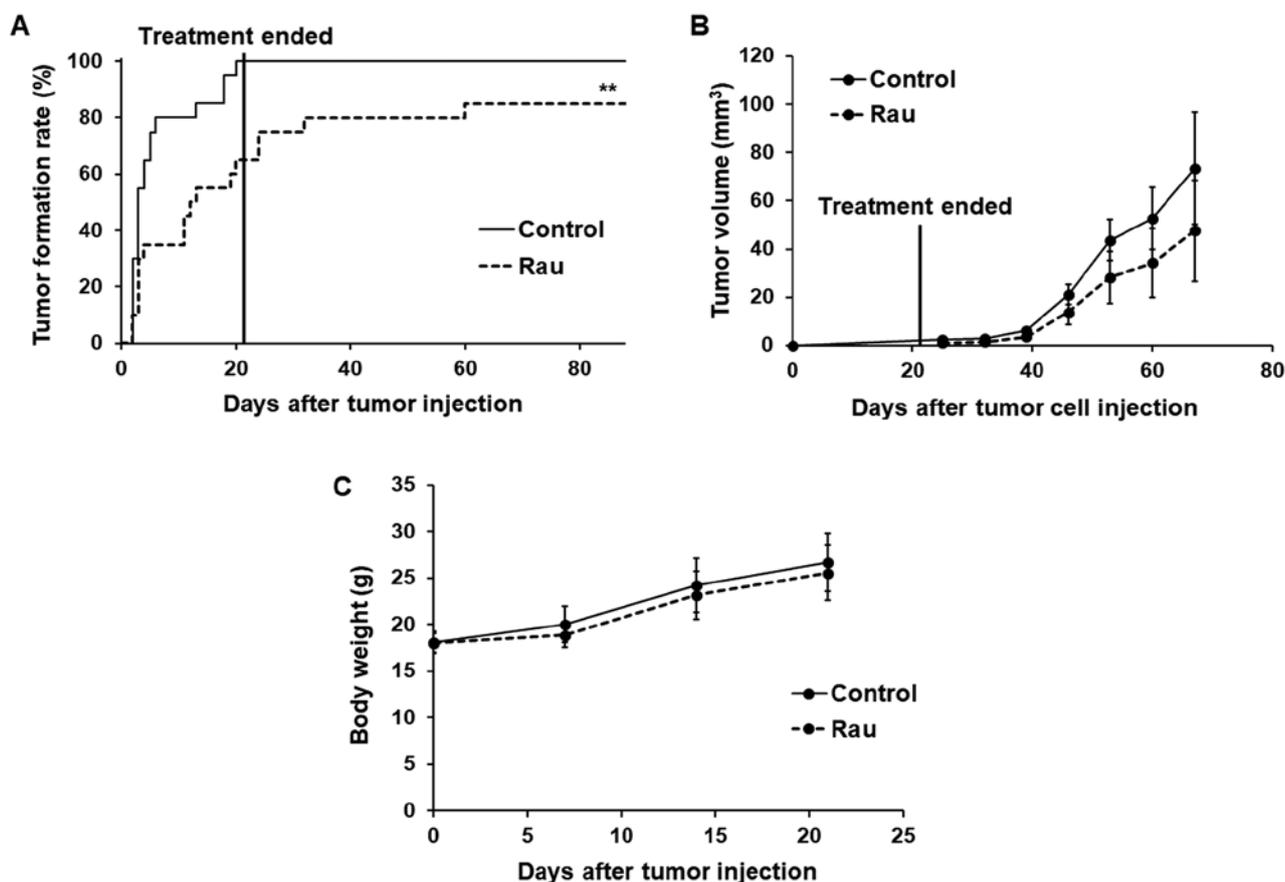


Figure 7. Effects of repeated Rau treatment on PANC-1 tumor formation and tumor growth in nude mice. (A) Tumor formation rate. 2×10^5 PANC-1 cells were mixed with 200 mg/ml Rau or PBS (control), and inoculated at both flanks of nude mice ($n=10$ for each group). Treatment started the next day and lasted for 3 weeks with oral gavage of 20 mg/kg Rau or saline (control) five times per week. $**P<0.01$ by log-rank test. (B) Longitudinal tumor growth. Tumor size was measured weekly using calipers. Tumor volume = width x width x length/2. (C) Body weight of mice during treatment for 3 weeks. Rau, *Rauwolfia vomitoria*.

treatment was performed with oral administration of Rau. The optimal cell number for injection was selected as 2×10^5 per injection. The mice ($n=10$) were injected subcutaneously in the left and right flanks with PANC-1 cells mixed with 200 mg/ml of Rau. Treatment started the following day and lasted for 3 weeks with oral gavage of 20 mg/kg Rau, five times per week. The control mice ($n=10$) were inoculated with the same number of cells mixed with PBS, and were administered with equivalent volumes of saline.

The rate of tumor formation and time of tumor formation were significantly different between the control and treated groups (Fig. 7A). At day 6, the tumor formation rate in the control group reached 80%, whereas that in the Rau-treated group was only 35%. At day 20 when the treatment had stopped, all mice in the control group were bearing tumors on both flanks (100% tumor formation), whereas the Rau-treated group only had 65% tumor formation. All mice were kept for 2 months following the end of treatment. At the end of the experiment, the Rau-treated group had a maximum of 85% tumor formation, compared with 100% tumor formation in the control group. These data indicated that Rau administration at 20 mg/kg orally eliminated CSCs in 15% of the injection sites.

The growth of the formed tumors was not significantly inhibited by Rau treatment compared with the control group, which indicated the lack of a long-term inhibitory effect on tumor growth following the end of treatment (Fig. 7B). No

adverse effects were observed in either group during the treatment (Fig. 7C).

Discussion

Targeting CSCs has been an attractive strategy for developing novel treatments with the aim of eliminating the entire cancer cell population. However, targeting CSCs has been challenging. First, as CSCs are only a small population in the bulk of cancer cells, anticancer agents that have cytotoxicity to the bulk of cancer cells do not necessarily inhibit CSCs (5,6). CSCs possess self-renewal ability and are able to give rise to new tumors (17). CSCs are also found to be drug resistant (12,41). The mechanism by which CSCs become drug resistant remains to be fully elucidated. A partial reason is the quiescent status of CSCs in a growing tumor. Other potential mechanisms are the upregulated expression of the ABCG2 transporter, which facilitates the efflux of chemotherapeutic drugs from the cytosol (41), overexpression of detoxifying enzymes, enhanced DNA repair ability, and overexpression of anti-apoptotic proteins (12). Given the roles of CSC in tumor generation, metastasis and drug resistance, the identification and development of novel drugs that can inhibit CSCs may lead to a promising outcome in the comprehensive inhibition of tumor growth, metastasis and recurrence, and overcoming drug resistance. In the present study, it was demonstrated that

the Rau extract inhibited pancreatic CSCs *in vitro* and *in vivo*. Previously, it was reported that the same Rau extract induced the apoptosis of pancreatic cancer cells and sensitized pancreatic cancer cells to gemcitabine treatment (29). The inhibition of CSCs may be another factor contributing to Rau-induced gemcitabine sensitivity in addition to its apoptosis-inducing activity. The data suggested that Rau had preferential inhibitory effects towards pancreatic CSCs, and also inhibited the bulk of cancer cells. This may be advantageous in cancer therapy as one treatment inhibits CSC and non-CSCs. The overall inhibitory effect in non-CSC and CSCs results in the inhibition of tumor growth, whereas the inhibition in CSCs is likely also to reduce metastasis and chemotherapy resistance. Given the lack of treatment options for pancreatic cancer, the benefits of Rau in pancreatic cancer treatment warrant further investigation, particularly in combination with current chemotherapies.

Dye exclusion and CSC surface markers are used in CSC isolation and provide consistent data for the enrichment of pancreatic CSCs. However, there has not been an efficient method to definitively identify and isolate a pure pancreatic CSCs and maintain/amplify them for drug development purposes (42). Functional assays, including tumor spheroid assays and tumorigenicity in mice are commonly used (43). Due to the difficulties in obtaining and maintaining a pure CSC population (44), the isolated CSCs may lose their natural environment in the bulk population (17). In the present study, the bulk of pancreatic cancer cells were treated, in addition to a dye-excluding side population, and the CSC-specific outcomes were examined. The inhibition of CSCs was shown, and was not likely due to the general cytotoxicity of Rau to the bulk of cancer cells. The data showed that Rau had an IC_{50} value of 317 $\mu\text{g/ml}$ over 48 h of treatment towards the bulk of PANC-1 cells, and had markedly lower IC_{50} values of 126.9-142.04 $\mu\text{g/ml}$ for the reduction of CD24⁺EpCam⁺ cells at a shorter treatment time of 24-48 h. Furthermore, in the tumor spheroid formation assay, Rau had an IC_{50} value of 39.44 $\mu\text{g/ml}$ in inhibiting the number of spheroids. These data suggested that Rau had a preferential inhibitory activity towards pancreatic CSCs.

The data obtained in the present study showed that Rau reduced protein levels of nuclear β -catenin and Nanog in PANC-1 cells, which are important in stem cell initiation and maintenance. Rau also reduced the mRNA levels of several CSC-related genes, namely Dppa4, Esrrb and Sox2. The in-depth mechanism underlying how Rau interacts with Nanog and/or the β -catenin signaling pathway requires further investigation. In addition, as plant extracts contain a complex mixture of natural compounds, it is possible that Rau also affects other molecular targets and pathways that lead to its CSC inhibitory effect.

Previous studies on extracts of Rau showed inhibitory effects on the proliferation on pancreatic, ovarian and prostate cancer (25,29,45). The data from animal experiments in the present study revealed the promising effects of Rau in inhibiting tumorigenicity, at a dose and administration route that can be easily translated into clinical use. No toxic side-effects were observed in mice at this dosage. The inhibition of tumorigenicity indicated the possible role of Rau in the prevention of cancer, in addition to data indicating a treatment

role. As extracts of *Rauwolfia vomitoria* are consumed by the American public as a health supplement, the safety, toxicity and effects of Rau as an anticancer agent require further investigation clinically.

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Availability of data and materials

All the original data concerning this publication is available upon request.

Authors' contributions

QC conceptualized, designed and oversaw the studies. RD participated in the study design and performed the majority of the experiments. PC performed part of the cell experiments. RD and PC collected the data. RD, PC and QC analyzed the data. All authors participated in manuscript preparation and all approved the manuscript and agree to be accountable for all aspects of the research in ensuring that the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Ethics approval and consent to participate

All animal experiments followed a protocol approved by the Institutional Animal Care and Use Committee of the University of Kansas Medical Center.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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